

## Review

# Bioactive lipids in atherosclerosis

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**ABSTRACT** Atherosclerosis is a low-grade systemic inflammatory condition and a dynamic process. Recent evidences suggest that anti-inflammatory products of polyunsaturated fatty acids such as lipoxins, resolvins, maresins and nitrolipids play a significant role in atherosclerosis by modulating the functions of platelets, leukocytes and macrophages and by protecting endothelial cells from the actions of reactive oxygen species. Hence, methods designed to enhance the formation of these bioactive anti-inflammatory lipids could be employed in the prevention and management of atherosclerosis.

**Key-words:** Atherosclerosis, polyunsaturated fatty acids, lipoxins, resolvins, protectins, free radicals, plaque, inflammation, nitrolipids.

## INTRODUCTION

Atherosclerosis, the major underlying cause for coronary heart disease (CHD), is a dynamic process. In majority of the instances, hyperlipidemia, diabetes mellitus, hypertension, obesity, hyperhomocysteinemia and smoking are the main risk factors for the development of atherosclerosis and CHD, conditions in which EFA (essential fatty acids) metabolism is abnormal such that plasma and tissue concentrations of  $\gamma$ -linolenic acid (GLA), dihomo-GLA (DGLA), arachidonic acid (AA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA) in the phospholipid fraction are low.<sup>1-8</sup> Increased intake of polyunsaturated fatty acids (PUFAs especially in the form of GLA, DGLA, EPA and DHA) protects against the development of these diseases both in experimental animals<sup>9-12</sup> and humans,<sup>13</sup> though the exact mechanism of this protective action is unclear. GLA, DGLA, AA, EPA, and DHA form precursors to prostaglandin E<sub>1</sub> (PGE<sub>1</sub>),

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prostacyclin (PGI<sub>2</sub>), PGI<sub>3</sub>, lipoxins (LXs), resolvins, neuroprotectin D1 (NPD1), enhance NO generation, and interact with NO to form nitrolipids that have anti-inflammatory actions, prevent platelet aggregation, inhibit leukocyte activation and augment wound healing and resolve inflammation that may account for their beneficial actions. This implies that an altered EFA metabolism in the form of a block in the activity of  $\Delta^6$  and  $\Delta^5$  desaturases, which are essential for the formation of long-chain metabolites from dietary linoleic acid (LA, 18:2  $\omega$ -6) and  $\alpha$ -linolenic acid (ALA, 18:3  $\omega$ -3), and inadequate formation of anti-inflammatory lipoxins, resolvins, protectins, maresins and nitrolipids from their precursor PUFAs could lead to the initiation, progression and aggravation of atherosclerosis.

### **LIPOXINS ARE POTENT ANTI-INFLAMMATORY MOLECULES**

Lipoxins and their aspirin-triggered carbon-15 epimers are key mediators of endogenous anti-inflammation and resolution. Aspirin-triggered lipoxin A4 analog (ATL-1) have been shown to modulate reactive oxygen species (ROS) generation in endothelial cells. Pre-treatment of endothelial cells with ATL-1 completely blocked ROS production triggered by different agents, inhibited the phosphorylation and translocation of the cytoplasmic NAD(P)H oxidase subunit p47 (phox) to the cell membrane as well as NAD(P)H oxidase activity and impaired the redox-sensitive activation of the transcriptional factor NF- $\kappa$ B, suggesting that lipoxins play a protective role against the development and progression of atherosclerosis and various cardiovascular diseases in which endothelial dysfunction is known to exist.<sup>14</sup> These results are supported in experiments performed with apolipoprotein E-deficient mice with (a) global leukocyte 12/15-lipoxygenase deficiency, (b) normal enzyme expression, or (c) macrophage-specific 12/15-lipoxygenase overexpression in which it was noted that 12/15-lipoxygenase expression protected mice against atherosclerosis via its role in the biosynthesis of lipoxin A4, resolvin D1, and protectin D1. These lipid mediators showed potent agonist actions on macrophages and vascular endothelial cells that reduced the magnitude of the local inflammatory response suggesting that a failure of local resolution mechanisms may underlie the unremitting inflammation that fuels atherosclerosis.<sup>15</sup> The evidence that lipoxins, resolvins and protectins are anti-inflammatory compounds and pro-inflammatory

cytokines are elevated in atherosclerosis lends support to the belief that atherosclerosis is an inflammatory condition.

### **ATHEROSCLEROSIS IS A LOW-GRADE SYSTEMIC INFLAMMATORY CONDITION**

An increase in the plasma concentrations of C-reactive protein (CRP), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-6 (IL-6), myeloperoxidase (MPO), lipoprotein associated phospholipase A<sub>2</sub> (Lp-PLA<sub>2</sub>), and lipid peroxides occurs in atherosclerosis suggesting that it is a low-grade systemic inflammatory condition.<sup>16-24</sup>

In atherosclerosis, circulating endothelial nitric oxide (eNO) levels are low,<sup>25-27</sup> reactive oxygen species (ROS) will be high, and anti-oxidant content will be low especially in the endothelial cells at atherosclerosis-prone areas of the blood vessels.<sup>28-32</sup> The decrease in the production of eNO by endothelial cells may, in part, be due to enhanced levels of asymmetrical dimethylarginine (ADMA) that inhibits eNO generation that may lead to increased mortality due to cardiovascular diseases.<sup>33-36</sup> What is more significant is the observation that plasma ADMA concentrations were found to be positively related to internal carotid/bulb intimal-media thickness, suggesting that ADMA promotes subclinical atherosclerosis in a site-specific manner, with a greater proatherogenic influence at known vulnerable sites in the arterial tree.<sup>37</sup> In addition to ADMA, homocysteine also augments the formation of superoxide anion and reduces the synthesis and release of eNO.<sup>38,39</sup> Homocysteine markedly reduced the increase in haem oxygenase (HO) activity and HO-1 protein expression induced by sodium nitroprusside. High levels of homocysteine also abolished hypoxia-mediated HO-1 expression.<sup>40</sup>

It is noteworthy that NO reacts with PUFAs to yield their respective nitroalkene derivatives that can be detected in plasma. These nitroalkene derivatives, termed as nitrolipids, produce vascular relaxation, inhibit neutrophil degranulation and superoxide formation, and inhibit platelet activation.<sup>41-43</sup> Nitrolipids have endogenous PPAR- $\gamma$  ligand activity and release NO.<sup>43</sup> These actions of nitrolipids prevent platelet aggregation, thrombus formation and atherosclerosis, and prevent inflammation.

Thus, PUFAs and their metabolites such as eicosanoids, lipoxins, resolvins, protectins, maresins and nitrolipids; various pro- and anti-inflammatory

cytokines, free radicals, nitric oxide (including ADMA) and various antioxidants seem to play critical role in the pathobiology of atherosclerosis.

### PLATELETS, LEUKOCYTES AND ENDOTHELIAL CELLS IN ATHEROSCLEROSIS

The cross-talk among platelets, leukocytes and endothelial cells could determine the initiation and progression of atherosclerosis. For instance, under normal conditions, endothelial cells produce adequate amounts of PGE<sub>1</sub> from DGLA; PGI<sub>2</sub> from AA; LXs, resolvins, protectins and maresins from AA, EPA and DHA; formation of adequate amounts of nitrolipids such that the pro-inflammatory and pro-atherosclerotic events are successfully abrogated. Some of the pro-inflammatory and pro-atherogenic stimuli include: hemodynamic forces, hyperlipidemia, hypertension, hyperglycemia, smoking, etc; that induce the expression of pro-inflammatory genes, which initiate and accelerate atherosclerosis at the points of shear stress. These factors enhance infiltration of intima by leukocytes and macrophages, cause low-level activation of NF- $\kappa$ B and elevated expression of VCAM-1 and ICAM-1, IL-1, IL-6, MCP-1, as well as antioxidant genes glutathione peroxidase and glutathione-S-transferase 2, and pro-inflammatory eicosanoids such as TXA<sub>2</sub>, PGE<sub>2</sub>, PGF<sub>2 $\alpha$</sub> , LTs, and other PGs, TXs, and LTs, and increased production and release of free radicals and UCP (uncoupling proteins) expression occurs in endothelial cells, platelets, and leukocytes in atherosclerosis-susceptible regions, and endothelial cells themselves may show changes in cell shape and proliferation. These adverse events can be prevented and atherosclerosis process is arrested by the production of adequate amounts of PGE<sub>1</sub>, PGI<sub>2</sub>, PGI<sub>3</sub>, LXs, resolvins, protectins, maresins, nitrolipids, NO, and anti-inflammatory cytokines such as IL-4, IL-10, TGF- $\beta$  by endothelial cells. Thus, the balance between pro- and anti-inflammatory and pro and anti-atherosclerotic factors is tilted more towards pro-atherosclerotic and pro-inflammatory factors, atherosclerosis occurs.<sup>44</sup>

### UNCOUPLING PROTEIN-1, ESSENTIAL FATTY ACIDS, AND ATHEROSCLEROSIS

The patchy manner in which atherosclerosis occurs suggests that arterial walls undergo regional disturbances of metabolism that include

the uncoupling of respiration and oxidative phosphorylation, which may be characteristic of blood vessels being predisposed to the development of atherosclerosis.<sup>45</sup> Oxidative stress is implicated in atherosclerosis. Mitochondrial electron transport accounts for most reactive oxygen species (ROS) production.<sup>46</sup> Uncoupling proteins (inner mitochondrial membrane anion transporters) allow protons to leak back into the mitochondrial matrix, thereby decreasing the potential energy available for ADP phosphorylation and ROS generation. Superoxide anion activates uncoupling proteins<sup>47,48</sup> that, in turn, limit further superoxide generation by dissipating proton motive force and thus, decreases oxidative stress. Uncoupling decreases glucose-induced ROS formation and abrogates pathways associated with vascular damage in endothelial cells *in vitro*.<sup>49</sup> In contrast, UCP-2 in macrophages decreases ROS and atherosclerosis.<sup>50</sup> Although, these results appear to be in conflict with the proposal that inefficient vascular metabolism is detrimental, it is known that uncoupling agents produce smooth muscle contraction and cause hypertension,<sup>51</sup> and it was reported that respiratory uncoupling is increased in the aortae of experimental animals that are susceptible to atherosclerosis.<sup>45</sup> These results imply that the efficiency of vascular wall energy metabolism could be a determinant of atherosclerotic lesion development. UCP-1 expression in aortic smooth muscle cells causes hypertension and increases atherosclerosis without affecting cholesterol levels.<sup>46,52</sup> This increase in UCP-1 expression also enhanced superoxide anion production and decreased the availability of NO, suggesting that oxidative stress has been elevated. This implies that inefficient metabolism in blood vessels causes atherosclerosis.

One of the earliest signs of atherosclerosis is the development of abnormal mitochondria in smooth muscle cells,<sup>53</sup> suggesting that mitochondrial dysfunction triggers the disease. Arteries have marginal oxygenation<sup>54</sup> and hypoxia reduces the respiratory control ratio.<sup>55</sup> Uncoupled respiration precedes atherosclerosis at lesion-prone sites but not at the sites that are resistant to atherosclerosis.<sup>45</sup> Disease-free aortae have abundant concentrations of the essential fatty acid-linoleate, whereas fatty streaks (an early stage of atherosclerosis) are deficient in EFAs.<sup>52,56,57</sup> EFA deficiency promotes respiratory uncoupling<sup>58,59</sup> and atherosclerosis.<sup>1,60,61</sup> Oxidative stress increases ROS generation and decreases NO formation and/or availability to be associated with

smooth muscle expression of UCP-1. These results emphasize that local disturbances of metabolism in the arterial wall are responsible for atherosclerosis and vascular disease.

## PUFAs IN ATHEROSCLEROSIS

Atherosclerotic plaque rupture is known to be responsible for sudden coronary events. Felton et al<sup>62</sup> reported that the concentrations of all fatty acids were increased at the edge of disrupted plaques compared with the center, but as a proportion of total fatty acids,  $\omega$ -6 were lower, suggesting that  $\omega$ -6 fatty acids have a significant role in atherosclerosis. It is possible that there is a close interaction between  $\omega$ -3 and  $\omega$ -6 fatty acids, which could influence one's susceptibility or resistance to atherosclerosis. It is interesting to note that EPA/DHA readily get incorporated into the atheromatous plaque, and patients treated with fish oil had more thick fibrous caps and no signs of inflammation compared with plaques in patients in the control and sunflower oil groups. Furthermore, the number of macrophages in plaques from patients receiving fish oil was lower than in the other two groups, suggesting that atherosclerotic plaques readily incorporate  $\omega$ -3 PUFAs from fish-oil supplementation, inducing changes that can enhance stability of atherosclerotic plaques.<sup>63</sup> In contrast, trans-fatty acids may render atheromatous plaques unstable, partly by displacing  $\omega$ -3 fatty acids, interfering with  $\omega$ -3 fatty acid metabolism and activating inflammatory responses and endothelial dysfunction.<sup>64,65</sup>

In this context, the interaction between  $\omega$ -3 and  $\omega$ -6 fatty acids is particularly significant. DGLA increases the conversion of EPA to PGI<sub>3</sub>, AA augmented the conversion of EPA to PGI<sub>3</sub>, EPA enhances the tissue levels of DGLA leading to increase in the formation of PGE<sub>1</sub>, events that prevent atherosclerosis. In contrast, trans-fats interfere with the formation of DGLA, AA, EPA, and DHA and thus, prevent the formation of anti-atherosclerotic molecules: PGE<sub>1</sub>, PGI<sub>2</sub>, PGI<sub>3</sub>, lipoxins, resolvins, protectins, maresins and nitrolipids and at the same time may augment the formation and/or action of LTs, and TXs that promote atherosclerosis. The beneficial action of statins (HMG-CoA reductase inhibitors) and glitazones (PPARs agonists) seem to be mediated by EFAs and their metabolites such as LXs, resolvins, and protectins,<sup>66-72</sup> which are potent anti-inflammatory molecules.<sup>1,73-75</sup> On the other hand, cholesterol and saturated fatty acids similar to trans-

fats block the activities of both  $\Delta$ 6 and  $\Delta$ 5 desaturases and inhibit the conversion of dietary LA and ALA to their respective long-chain metabolites and render cell membrane more rigid.<sup>1</sup> Increase in the consumption of trans-fats, cholesterol, and saturated fatty acids enhanced,<sup>76-78</sup> whereas consumption of  $\omega$ -3 fatty acids decreased the levels of inflammatory markers.<sup>79</sup> These evidences suggest that  $\omega$ -3 and  $\omega$ -6 fatty acids, trans-fats, saturated fatty acids and cholesterol modulate inflammation and thus, influence the pathobiology of atherosclerosis, CAD and stroke.

## ATHEROPROTECTIVE ACTIONS OF $\omega$ -3 AND $\omega$ -6 FATTY ACIDS

It is evident from the preceding discussion that both  $\omega$ -3 and  $\omega$ -6 PUFAs interact with each other to prevent atherosclerosis, CAD, CVD, and stroke, though  $\omega$ -3 EPA and DHA seem to be having a more dominant role compared to  $\omega$ -6 in this beneficial action. PUFAs display a multitude of actions (such as ability to lower plasma triglycerides, cholesterol and apolipoprotein B and alter hemostatic system; see table 1 also for the actions of PUFAs on lipid metabolism) to prevent atherosclerosis.

## CONCLUSIONS

Based on the preceding discussion, it is clear that atherosclerosis is a low-grade inflammatory condition and PUFAs (especially  $\omega$ -3 EPA and DHA) are useful in its prevention and management. PUFAs also inhibit ACE and HMG-CoA reductase activities and behave as endogenous ACE inhibitors. Statins similar to PUFAs and their products such as lipoxins, resolvins, protectins,

**TABLE 1.** Summary of effects of PUFAs on nuclear receptors involved in the regulation of lipogenesis.

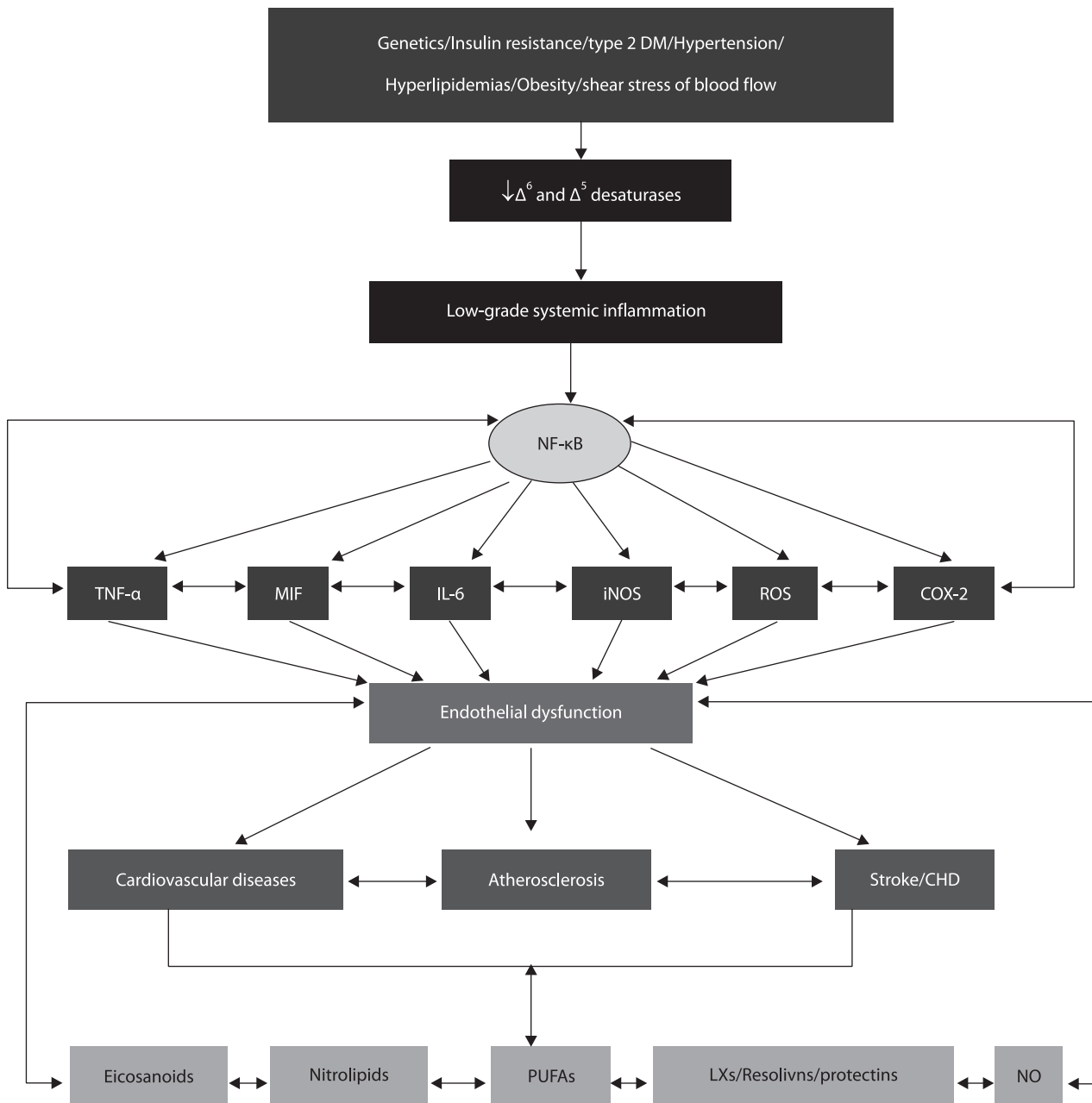
Nuclear receptor	Effects on gene regulation	Expected changes		
		TG	HDL	LDL
PPAR- $\alpha$	↑	↓↓	↑	↓
LXR	↓	↓↓	↓	↓
FXR	↑	↓↓	↑	↑
HNF-4 $\alpha$	↓	↓↓	↓	↔

FXR=Farnesol X receptor; HDL=High-density lipoprotein; HNF-4 $\alpha$ =Hepatocyte nuclear factor-4 $\alpha$ ; LDL=Low-density lipoprotein, LXR=Liver X receptor; PPAR- $\alpha$ =Peroxisome proliferator-activated receptor; ↑=Increase; ↓=Decrease; ↔=Neutral effect

maresins and nitrolipids suppress the production of pro-inflammatory cytokines, modulate SREBP pathway and thus, inhibit atherosclerosis both by lowering plasma triglycerides and cholesterol levels (see table 1), and modulating inflammatory events.

These evidences suggest that atherosclerosis can be prevented/arrested if endothelial cells are able to

produce adequate amounts of various PUFAs such that they in turn lead to the formation of beneficial PGE1, PGI2, PGI3, LXs, resolvins, protectins, maresins and nitrolipids that are capable of suppressing inflammation, expression of various adhesion molecules on the surface of endothelial cells, and prevent leukocyte, monocyte and macrophage infiltration of endothelial cells (see figure 1).



**FIGURE 1.** Scheme showing the relationship among various mediators of endothelial dysfunction and CHD/stroke and the role of PUFAs and their metabolites in these processes.

## Βιοενεργά λιπίδια στην αθηροσκλήρωση

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**ΠΕΡΙΛΗΨΗ** Η αθηροσκλήρωση αποτελεί μια χαμηλού βαθμού συστηματική φλεγμονώδη κατάσταση και συγχρόνως μια δυναμική διαδικασία. Πρόσφατα δεδομένα υποστηρίζουν ότι τα αντιφλεγμονώδη παράγωγα των πολυακόρεστων λιπαρών οξέων, όπως οι λιποξίνες, οι ρεσολβίνες, οι μαρεσίνες και τα νιτρολιπίδια, διαδραματίζουν σημαντικό ρόλο στη διαδικασία της αθηροσκλήρωσης, τροποποιώντας τις λειτουργίες των αιμοπεταλίων, των λευκοκυττάρων και των μακροφάγων και προστατεύοντας τα ενδοθηλιακά κύτταρα από τις δράσεις των ενεργών μορφών οξυγόνου. Ως εκ τούτου, προσεγγίσεις που αποσκοπούν στην ενίσχυση του σχηματισμού αυτών των βιοενεργών αντιφλεγμονωδών λιπιδίων θα μπορούσαν να ενταχθούν στην πρόληψη και αντιμετώπιση της αθηροσκλήρωσης.

**Λέξεις ευρητηρίου:** Αθηροσκλήρωση, πολυακόρεστα λιπαρά οξέα, λιποξίνες, ρεσολβίνες, προτεκτίνες, ελεύθερες ρίζες, πλάκα, φλεγμονή, νιτρολιπίδια.

### References

1. Das UN. Essential fatty acids: Biochemistry, physiology and pathology. *Biotech J* 2006, 1:420-439
2. Das UN. Essential fatty acid metabolism in patients with essential hypertension, diabetes mellitus and coronary heart disease. *Prostagland Leukotr Essent Fatty Acids* 1995, 52:387-391
3. Kumar KV, Das UN. Lipid peroxides and essential fatty acids in patients with coronary heart disease. *J Nutritional Med* 1994, 4:33-37
4. Das UN. Nutritional factors in the pathobiology of human essential hypertension. *Nutrition* 2001, 17:337-346
5. Das UN. Can perinatal supplementation of long chain polyunsaturated fatty acids prevent hypertension in adult life? *Hypertension* 2001, 38:e6-e8
6. Das UN. Can perinatal supplementation of long-chain polyunsaturated fatty acids prevent diabetes mellitus? *Eur J Clin Nutr* 2003, 57:218-226
7. Das UN. A defect in the activity of D6 and D5 desaturases may be a factor predisposing to the development of insulin resistance syndrome. *Prostagland Leukotr Essent Fatty Acids* 2005, 72:343-350
8. Wang L, Folsom AR, Eckfeldt JH. Plasma fatty acid composition and incidence of coronary heart disease in middle aged adults: the Atherosclerosis Risk in Communities (ARIC) Study. *Nutr Metab Cardiovasc Dis* 2003, 13:256-266
9. Zheng ZJ, Folsom AR, Ma J, Arnett DK, McGovern PG, Eckfeldt JH. Plasma fatty acid composition and 6-year incidence of hypertension in middle-aged adults: the Atherosclerosis Risk in Communities (ARIC) Study. *Am J Epidemiol* 1999, 150:492-500
10. Suresh Y, Das UN. Differential effect of saturated, monounsaturated, and polyunsaturated fatty acids on alloxan-induced diabetes mellitus. *Prostagland Leukotr Essential Fatty Acids* 2006, 74:199-213
11. Suresh Y, Das UN. Long-chain polyunsaturated fatty acids and chemically-induced diabetes mellitus: Effect of  $\omega$ -6 fatty acids. *Nutrition* 2003, 19:93-114
12. Suresh Y, Das UN. Long-chain polyunsaturated fatty acids and chemically-induced diabetes mellitus: Effect of  $\omega$ -3 fatty acids. *Nutrition* 2003, 19:213-228
13. Mozaffarian D, Ascherio A, Hu FB, Stampfer MJ, Willett WC, Siscovick DS, Rimm EB. Interplay between different polyunsaturated fatty acids and risk of coronary heart disease in men. *Circulation* 2005, 111:157-164
14. Nascimento-Silva V, Arruda MA, Barja-Fidalgo C, Fierro IM. Aspirin-triggered lipoxin A4 blocks reactive oxygen species generation in endothelial cells: a novel antioxidative mechanism. *Thromb Haemost* 2007, 97:88-98
15. Merched AJ, Ko K, Gotlinger KH, Serhan CN, Chan L. Atherosclerosis: evidence for impairment of resolution of vascular inflammation governed by specific lipid mediators. *FASEB J* 2008, 22:3595-3606
16. Das UN. Clinical laboratory tools to diagnose inflammation. *Adv Clin Chem* 2006, 41:189-229
17. Brennan ML, Penn MS, Lente FV, Nambi V, Shishehbor MH, Aviles RJ, Goormastic M, Pepoy ML, McErlan ES, Topol EJ, Nissen SE, Hazen SL. Prognostic value of myeloperoxidase in patients with chest pain. *N Engl J Med* 2003, 349:1595-1604
18. Ballantyne CM, Hoogeveen RC, Bang H, Coresh J, Folsom AR, Heiss G, Sharrett AR. Lipoprotein-associated phospholipase A2, high-sensitivity C-reactive protein, and risk for incident coronary heart disease in middle-aged men and women in the Atherosclerosis Risk in Communities (ARIC) Study. *Circulation* 2004, 109:837-842
19. Oei HHS, van der Meer IM, Hofman A, Koudstaal PJ, Stijnen T, Breteler MMB, Witteman JCM. Lipoprotein-associated phospholipase A2 activity is associated with risk of coronary heart disease and ischemic stroke. The Rotterdam study. *Circulation* 2005, 111:570-575
20. Häkkinen T, Luoma JS, Hiltunen MO, Macphee CH, Milliner KJ, Patel L, Rice SQ, Tew DG, Karkola K, Ylä-Herttuala S. Lipoprotein-associated phospholipase A(2), platelet-activating factor acetylhydrolase, is expressed by macrophages in human and rabbit atherosclerotic lesions. *Arterioscler Thromb Vasc Biol* 1999, 19:2909-2917
21. Lp-PLA(2) Studies Collaboration, Thompson A, Gao P, Orfei L, Watson S, Di Angelantonio E, Kaptoge S, Ballantyne C,

- Cannon CP, Criqui M, Cushman M, Hofman A, Packard C, Thompson SG, Collins R, Danesh J. Lipoprotein-associated phospholipase A(2) and risk of coronary disease, stroke, and mortality: collaborative analysis of 32 prospective studies. *Lancet* 2010, 375:1536–1544
22. Anuurad E, Ozturk Z, Enkhmaa B, Pearson TA, Berglund L. Association of lipoprotein-associated phospholipase A2 with coronary artery disease in African-Americans and Caucasians. *J Clin Endocrinol Metab* 2010, 95:2376–2383
  23. Hatoum IJ, Hu FB, Nelson JJ, Rimm EB. Lipoprotein-associated phospholipase A2 activity and incident coronary heart disease among men and women with type 2 diabetes. *Diabetes* 2010, 59:1239–1243
  24. Brilakis ES, Khera A, Saeed B, Banerjee S, McGuire DK, Murphy SA, de Lemos JA. Association of lipoprotein-associated phospholipase A2 mass and activity with coronary and aortic atherosclerosis: findings from the Dallas Heart Study. *Clin Chem* 2008, 54:1975–1981
  25. Wilcox JN, Subramanian RR, Sundell CL, Tracey WR, Pollock JS, Harrison DG, Marsden PA. Expression of multiple isoforms of nitric oxide synthase in normal and atherosclerotic vessels. *Arterioscler Thromb Vasc Biol* 1997, 17:2479–2488
  26. Garlich CD, Beyer J, Zhang H, Schmeisser A, Plötze K, Mügge A, Schellong S, Daniel WG. Decreased plasma concentrations of L-hydroxy-arginine as a marker of reduced NO formation in patients with combined cardiovascular risk factors. *J Lab Clin Med* 2000, 135:419–425
  27. Napoli C, Ignarro LJ. Nitric oxide and atherosclerosis. *Nitric Oxide* 2001, 5:88–97
  28. Loffredo L, Pignatelli P, Cangemi R, Andreozzi P, Panico MA, Meloni V, Violi F. Imbalance between nitric oxide generation and oxidative stress in patients with peripheral arterial disease: effect of an antioxidant treatment. *J Vasc Surg* 2006, 44:525–530
  29. Aygul R, Kotan D, Demirbas F, Ulvi H, Deniz O. Plasma oxidants and antioxidants in acute ischaemic stroke. *J Int Med Res* 2006, 34:413–418
  30. Collino M, Aragno M, Mastrocola R, Gallicchio M, Rosa AC, Dianzani C, Danni O, Thiemermann C, Fantozzi R. Modulation of the oxidative stress and inflammatory response by PPAR-gamma agonists in the hippocampus of rats exposed to cerebral ischemia/reperfusion. *Eur J Pharmacol* 2006, 530:70–80
  31. Kumar KV, Das UN. Lipid peroxides and essential fatty acids in patients with coronary heart disease. *J Nutritional Med* 1994, 4:33–37
  32. Das UN. Free radicals, cytokines and nitric oxide in cardiac failure and myocardial infarction. *Mol Cell Biochem* 2000, 215:145–152
  33. Zoccali C, Bode-Böger S, Mallamaci F, Benedetto F, Tripepi G, Malatino L, Cataliotti A, Bellanuova I, Fermo I, Frölich J, Böger R. Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: a prospective study. *Lancet* 2001, 358:2113–2117
  34. Maas R, Quitzau K, Schwedhelm E, Spiekler L, Rafflenbeul W, Steenpass A, Lüscher TF, Böger RH. Asymmetrical dimethylarginine (ADMA) and coronary endothelial function in patients with coronary artery disease and mild hypercholesterolemia. *Atherosclerosis* 2007, 191:211–219
  35. Juonala M, Viikari JS, Alftan G, Marniemi J, Kähönen M, Taittonen L, Laitinen T, Raitakari OT. Brachial artery flow-mediated dilation and asymmetrical dimethylarginine in the cardiovascular risk in young Finns study. *Circulation* 2007, 116:1367–1373
  36. Antoniadis C, Shirodaria C, Leeson P, Antonopoulos A, Warrick N, Van-Assche T, Cunnington C, Tousoulis D, Pillai R, Ratnatunga C, Stefanadis C, Channon KM. Association of plasma asymmetrical dimethylarginine (ADMA) with elevated vascular superoxide production and endothelial nitric oxide synthase uncoupling: implications for endothelial function in human atherosclerosis. *Eur Heart J* 2009, 30:1142–1150
  37. Maas R, Xanthakis V, Polak JF, Schwedhelm E, Sullivan LM, Benndorf R, Schulze F, Vasani RS, Wolf PA, Böger RH, Seshadri S. Association of the endogenous nitric oxide synthase inhibitor ADMA with carotid artery intimal media thickness in the Framingham Heart Study offspring cohort. *Stroke* 2009, 40:2715–2719
  38. Ungvari Z, Pacher P, Rischák K, Szollár L, Koller A. Dysfunction of nitric oxide mediation in isolated rat arterioles with methionine diet-induced hyperhomocysteinemia. *Arterioscler Thromb Vasc Biol* 1999, 19:1899–1904
  39. Das UN. Folic acid says NO to vascular diseases. *Nutrition* 2003, 19:686–692
  40. Sawle P, Foresti R, Green CJ, Motterlini R. Homocysteine attenuates endothelial haem oxygenase-1 induction by nitric oxide (NO) and hypoxia. *FEBS Lett* 2001, 508:403–406
  41. Baker PRS, Lin Y, Schopfer FJ, Woodcock SR, Groeger AL, Batthyany C, Sweeney S, Long MH, Iles KE, Baker LMS, Branchaud BP, Chen Y, Freeman BA. Fatty acid transduction of nitric oxide signaling: Multiple nitrated unsaturated fatty acid derivatives exist in human blood and urine and serve as endogenous peroxisome proliferator-activated receptor ligands. *J Biol Chem* 2005, 280:42464–42475
  42. Coles B, Bloodsworth A, Clark SR, Lewis MJ, Cross AR, Freeman BA, O'Donnell VB. Nitrolinoleate inhibits superoxide generation, degranulation, and integrin expression by human neutrophils. *Circ Res* 2002, 91:375–381
  43. Lima ES, Bonim MG, Augusto O, Barbeiro HV, Souza HP, Abdalla DSP. Nitrated lipids decompose to nitric oxide and lipid radicals and cause vasorelaxation. *Free Radic Biol Med* 2005, 39:532–539
  44. Das UN. Crosstalk among leukocytes, platelets, and endothelial cells and its relevance to atherosclerosis and coronary heart disease. *Current Nutrition Food Sci* 2009, 5:75–93
  45. Santerre RF, Nicolosi RJ, Smith SC. Respiratory control in preatherosclerotic susceptible and resistant pigeon aortas. *Exp Mol Pathol* 1974, 20:397–406
  46. Droge W. Free radicals in the physiological control of cell function. *Physiol Rev* 2002, 82:47–95
  47. Echtay KS, Roussel D, St-Pierre J, Jekabsons MB, Cadenas S, Stuart JA, Harper JA, Roebuck SJ, Morrison A, Pickering S, Clapham JC, Brand MD. Superoxide activates mitochondrial uncoupling proteins. *Nature* 2002, 415:96–99
  48. Murphy MP, Echtay KS, Blaikie FH, Asin-Cayuela J, Cocheme HM, Green K, Buckingham JA, Taylor ER, Hurrell F, Hughes G.

- Miwa S, Cooper CE, Svistunenko DA, Smith RA, Brand MD. Superoxide activates uncoupling proteins by generating carbon-centered radicals and initiating lipid peroxidation: studies using a mitochondria-targeted spin trap derived from alpha-phenyl-N-tert-butyl nitron. *J Biol Chem* 2003, 278:48534–48545
49. Nishikawa T, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, Yorek MA, Beebe D, Oates PJ, Hammes HP, Giardino I, Brownlee M. Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature* 2000, 404:787–790
  50. Blanc J, Alves-Guerra MC, Esposito B, Rousset S, Gourdy P, Rixquier D, Tedgui A, Miroux B, Mallat Z. Protective role of uncoupling protein 2 in atherosclerosis. *Circulation* 2003, 107:388–390
  51. Petterson G. Effect of dinitrophenol and anoxia on isometric tension in rabbit colon smooth muscle. *Acta Pharmacol Toxicol (Copenh)* 1985, 57:184–189
  52. Bernal-Mizrachi C, Gates AC, Weng S, Imamura T, Knutsen RH, DeSantis P, Coleman T, Townsend RR, Muglia LJ, Semenkovich CF. Vascular respiratory uncoupling increases blood pressure and atherosclerosis. *Nature* 2006, 435:502–506
  53. Watts H. In: Jones RJ (ed) *Evolution of the Atherosclerotic Plaque*, Univ Chicago, Chicago, 1963, 117
  54. Levin M, Leppanen O, Evaldsson M, Wiklund O, Bondjers G, Bjornheden T. Mapping of ATP, glucose, glycogen, and lactate concentrations within the arterial wall. *Arterioscler Thromb Vasc Biol* 2003, 25:1801–1807
  55. Jennings RB, Kaltenbach JP, Sommens HM. Mitochondrial metabolism in ischemic injury. *Arch Pathol* 1967, 84:15–19
  56. Smith EB. The effects of age and of early atheromata on the intimal lipids in men. *Biochem J* 1962, 84:49
  57. Smith EB. Lipids carried by S1 0-12 lipoprotein in normal and hypercholesterolaemic serum. *Lancet* 1962, 2:530–534
  58. Klein PD, Johnson RM. Phosphorous metabolism in unsaturated fatty acid-deficient rats. *J Biol Chem* 1954, 211:103–110
  59. Hayashida T, Portman OW. Swelling of liver mitochondria from rats fed diets deficient in essential fatty acids. *Proc Soc Exp Biol Med* 1960, 103:656–659
  60. Cornwell DG, Panganamala RV. Atherosclerosis an intracellular deficiency in essential fatty acids. *Prog Lipid Res* 1981, 20:365–376
  61. Das UN. Essential fatty acids—a review. *Current Pharmaceutical Biotech* 2006, 7:467–482
  62. Felton CV, Crook D, Davies MJ, Oliver MF. Relation of plaque lipid composition and morphology to the stability of human aortic plaques. *Arterioscler Thromb Vasc Biol* 1997, 17:1337–1345
  63. Thies F, Garry JM, Yaqoob P, Rerkasem K, Williams J, Shearman CP, Gallagher PJ, Calder PC, Grimble RF. Association of n-3 polyunsaturated fatty acids with stability of atherosclerotic plaques: a randomised controlled trial. *Lancet* 2003, 361:477–485
  64. Mozaffarian D. Trans fatty acids—effects on systemic inflammation and endothelial function. *Atheroscler* 2006, (Suppl)7:29–32
  65. Mozaffarian D, Rimm EB, King IB, Lawler RL, McDonald GB, Levy WC. Trans fatty acids and systemic inflammation in heart failure. *Am J Clin Nutr* 2004, 80:1521–1525
  66. Das UN. Essential fatty acids as possible mediators of the actions of statins. *Prostaglandin Leukotr Essent Fatty Acids* 2001, 65:37–40
  67. Levine L. Statins stimulate arachidonic acid release and prostaglandin I<sub>2</sub> production in rat liver cells. *Lipids Health Dis* 2003, 2:1
  68. Jula A, Marniemi J, Ronnema T, Virtanen A, Huupponen R. Effects of diet and simvastatin on fatty acid composition in hypercholesterolemic men: a randomized controlled trial. *Arterioscler Thromb Vasc Biol* 2005, 25:1952–1959
  69. Harris JJ, Hibbeln JR, Mackey RH, Muldoon MF. Statin treatment alters serum n-3 and n-6 fatty acids in hypercholesterolemic patients. *Prostaglandin Leukotr Essent Fatty Acids* 2004, 71:263–269
  70. Bellini MJ, Polo MP, de Alaniz MJ, de Bravo MG. Effect of simvastatin on the uptake and metabolic conversion of palmitic, dihomo-gamma-linoleic and alpha-linolenic acids in A549 cells. *Prostaglandin Leukotr Essent Fatty Acids* 2003, 69:351–367
  71. Rise P, Pazzucconi F, Sirtori CR, Galli C. Statins enhance arachidonic acid synthesis in hypercholesterolemic patients. *Nutr Metab Cardiovasc Dis* 2001, 11:88–94
  72. Birnbaum Y, Ye Y, Lin Y, Freeberg SY, Nishi SP, Martinez JD, Huang MH, Uretsky BF, Perez-Polo JR. Augmentation of myocardial production of 15-epi-lipoxin-A<sub>4</sub> by pioglitazone and atorvastatin in the rat. *Circulation* 2006, 114:929–935
  73. Morris T, Stables M, Gilroy DW. New perspectives on aspirin and the endogenous control of acute inflammatory resolution. *Scien World J* 2006, 6:1048–1065
  74. Schwab JM, Serhan CN. Lipoxins and new lipid mediators in the resolution of inflammation. *Curr Opin Pharmacol* 2006, 6:414–420
  75. Serhan CN. Novel omega-3-derived local mediators in anti-inflammation and resolution. *Pharmacol Ther* 2005, 105:7–21
  76. Mozaffarian D, Pischon T, Hankinson SE, Rifai N, Joshipura K, Willett WC, Rimm EB. Dietary intake of trans fatty acids and systemic inflammation in women. *Am J Clin Nutr* 2004, 79:606–612
  77. Baer DJ, Judd JT, Clevidence BA, Tracy RP. Dietary fatty acids affect plasma markers of inflammation in healthy men fed controlled diets: a randomized crossover study. *Am J Clin Nutr* 2004, 79:969–973
  78. Lopez-Garcia E, Schulze MB, Meigs JB, Manson JE, Rifai N, Stampfer MJ, Willett WC, Hu FB. Consumption of trans fatty acids is related to plasma biomarkers of inflammation and endothelial dysfunction. *J Nutr* 2005, 135:562–566
  79. Lopez-Garcia E, Schulze MB, Manson JE, Meigs JB, Albert CM, Rifai N, Willett WC, Hu FB. Consumption of (n-3) fatty acids is related to plasma biomarkers of inflammation and endothelial activation in women. *J Nutr* 2004, 134:1806–1811